



Review

Anti-inflammatory and side effects of cyclooxygenase inhibitors

Halis Süleyman¹, Berna Demircan², Yalçın Karagöz³

¹Department of Pharmacology, ²Department of Biochemistry, Faculty of Medicine, Atatürk University, TR-25240, Erzurum, Turkey

³Graduate School of Natural and Applied Sciences, TR-25240, Erzurum, Turkey

Correspondence: Halis Süleyman, e-mail: suleyman@atauni.edu.tr

Abstract:

Nonsteroidal anti-inflammatory drugs (NSAIDs) are the most commonly used drugs in inflammatory diseases, since they are effective in management of pain, fever, redness, edema arising as a consequence of inflammatory mediator release. Studies have shown that both therapeutic and side effects of NSAIDs are dependent on cyclooxygenase (COX) inhibition. COX isoforms have been named constitutive (COX-1) and inducible (COX-2). COX-1 catalyzes formation of cytoprotective prostaglandins in thrombocytes, vascular endothelium, stomach mucosa, kidneys, pancreas, Langerhans islets, seminal vesicles, and brain. Induction of COX-2 by various growth factors, proinflammatory agents, endotoxins, mitogens, and tumor agents indicates that this isoform may have a role in induction of pathological processes, such as inflammation. It is well known that therapy with COX inhibitors is associated with a number of side effects including gastrointestinal erosions, and renal and hepatic insufficiency. Such critical adverse reactions are mostly dependent on COX-1 inhibition. As a result of research focused on reduction of the adverse effects of NSAIDs, selective COX-2 inhibitors, such as celecoxib and rofecoxib have been developed. However, many data demonstrate that mechanisms of action of these drugs are multidirectional and complex. These drugs or their derivatives, which belong to the same group, have distinct pharmacological effects, side effects and potencies which implies that there may be more than two, five or even tens of COX isoforms.

Key words:

NSAIDs, COX enzymes, inflammation, COX-2 selective inhibitors, non-selective inhibitors

Introduction

Nonsteroidal anti-inflammatory drugs (NSAIDs) are the most commonly used drugs in inflammatory diseases, since they are effective in management of pain, fever, redness, edema arising as a consequence of inflammatory mediator release [22, 57]. Studies have shown that both therapeutic and side effects of NSAIDs are dependent on cyclooxygenase (COX) in-

hibition [101]. It has been suggested that COX-2 inhibition is responsible for the therapeutic effects of NSAIDs, while COX-1 inhibition causes the gastrointestinal and renal side effects [60, 106, 107].

In this review, the anti-inflammatory and side effects of nonselective COX inhibitors and selective COX-2 inhibitors will be discussed and compared. COX isoenzymes, which form the basis of the mechanism of action of NSAIDs, will be described in order to make an understanding of the subject easier.

COX isoenzymes

Until the beginning of the 1990s, it had been thought that there was only one COX enzyme. In 1990, rapid development of the studies in this field revealed that COX enzyme had two distinct isoforms with different genetic coding [42, 75]. Although both isoforms had similar amino acid sequence and catalytic activity, they were demonstrated to have different functions [49, 106]. These isoforms were named ‘constitutive’ COX-1 and ‘inducible’ COX-2. COX-1 catalyzes the formation of cytoprotective prostaglandins (PGs) in thrombocytes, vascular endothelium, stomach mucosa, kidneys, pancreas, Langerhans islets, seminal vesicles, and brain [36, 57, 90]. Figure 1 gives a summary of the biosynthesis of PGs: the first step in the biosynthesis of prostanoids catalyzed by phospholipase A_2 is arachidonic acid (AA) release from the membrane phospholipids. The second step is AA conversion by cyclooxygenase. First, the unstable PGG_2 is produced in the COX reaction, which is then immediately converted into PGH_2 by the same enzyme in a peroxidase reaction. The end products of the AA metabolism are PGs, thromboxanes and prostacyclin [18]. Induction of COX-2 by various growth factors, proinflammatory agents, endotoxins, mitogens, tumor

agents [21, 54, 56] indicates that this isoform may have a role in formation of pathological processes, such as inflammation [73, 96]. COX-1 products, prostaglandins (PGI_2 and PGE_2), maintain integrity of gastrointestinal system (GIS) by reducing gastric acid

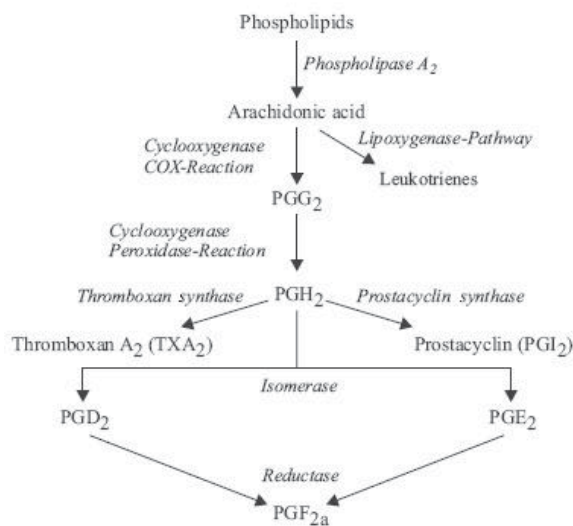


Fig. 1. Arachidonic acid cascade (Reprinted with permission of Elsevier Ltd., [18])

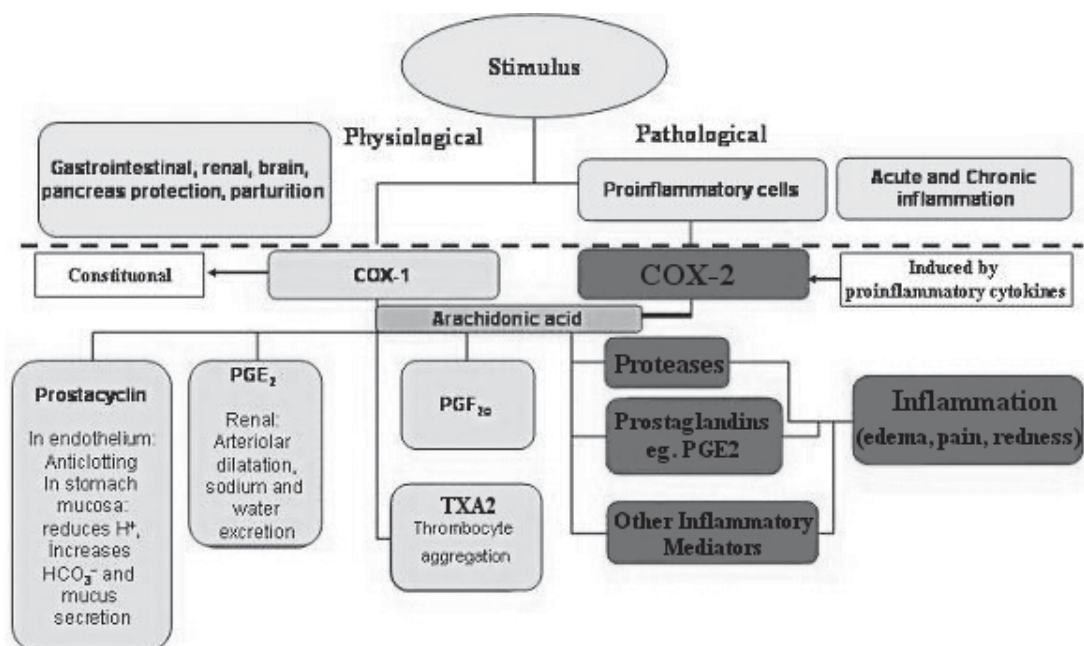


Fig. 2. Roles of COX-1 and COX-2 [18]

secretion, increasing the thickness of mucus layer, stimulating bicarbonate secretion and enhancing mucosal blood flow [26, 32, 86, 90]. PGE₂ enhances mucus secretion by activating cAMP in gastric epithelial cells (Fig. 2) [87]. Glucocorticoids and endogenous steroids can suppress the gene responsible for COX-2 synthesis [50, 56]. Drugs, which inhibit COX-1 more than COX-2, such as indomethacin, naproxen, ibuprofen, cause more severe damage to the gastric tissues [43]. As a result of studies focused on reduction of the adverse effects of NSAIDs, selective COX-2 inhibitors, such as celecoxib and rofecoxib, have been developed. Today, it is a well-known hypothesis in medicine that COX-1 is constitutive and cytoprotective, while COX-2 is an inducible enzyme in the inflamed tissues (Fig. 2). However, recent studies [59, 100] have questioned this hypothesis. Continuing research on COX isoforms has indicated that paracetamol has a more potent effect on COX preparations from the brain than on COX preparations from the spleen. It has been proposed that the third isoform of the enzyme, COX-3, may exist in the brain [23, 94].

Classification of NSAIDs

NSAIDs can be grouped in four categories:

1. Selective COX-1 inhibitors, such as aspirin;
2. Non-selective COX inhibitors: a number of the NSAIDs examined exhibit COX-1/COX-2 IC₅₀ ratios between 0.5 and 3.0;
3. Relatively selective COX-2 inhibitors, such as meloxicam, nimesulide, diclofenac with COX-1/COX-2 IC₅₀ ratios of 10–20;
4. Highly selective COX-2 inhibitors comprising three experimental compounds (NS-398, L-745, 337 and SC-58125, the prototype of celecoxib) with COX-1/COX-2 IC₅₀ ratios from 140 to 250 and rofecoxib with the ratio > 400 (Tab. 1) [60].

General information about NSAIDs

These drugs are named NSAIDs because they are structurally different from steroidal anti-inflammatory drugs [95]. As NSAIDs possess analgesic, antipyretic

Tab. 1. COX isoform selectivity assessed in the whole blood assays *in vitro* by cyclooxygenase inhibitors (Reprinted with permission of Elsevier Ltd., [60])

Inhibitor	COX-1/COX-2 IC ₅₀ ratio	
Aspirin	0.01	Selective COX-1 inhibitors
S-Indobufen	0.043	
Valeryl Salicylate	< 0.24	
Ibuprofen	0.50	Non-selective COX inhibitors
Naproxen	0.56	
S-Ketoprofen	0.61	
Flurbiprofen	1.00	
Sodium Salicylate	1.03	
6-MNA ^a	1.49	
Indomethacin	1.90	
Piroxicam	3.12	Relatively selective COX-2 inhibitors
Meloxicam	11.16	
Nimesulide	17.69	
Diclofenac	18.90	Highly selective COX-2 inhibitors
SC-58125	143.30	
NS-398	168.00	
L-745,337	246.00	
Rofecoxib	410.00	

^a 6-MNA is the active metabolite of nabumetone

and anti-inflammatory effects, they have certain advantages in the treatment of diseases with pain, fever and inflammation, when compared to steroidal anti-inflammatory and narcotic analgesic drugs [2]. Although NSAIDs have distinct chemical structures, they have similar therapeutic and side effects [64].

Mechanisms of action of NSAIDs

These mechanisms comprise inhibition of synthesis of COX and leukotriene (LO) products, prevention of release of toxic oxygen radicals and lysosomal enzymes, prevention of neutrophil aggregation, adhesion and chemotaxis, and uncoupling of oxidative phosphorylation [4, 10, 14, 29, 33, 48]. Recently, it

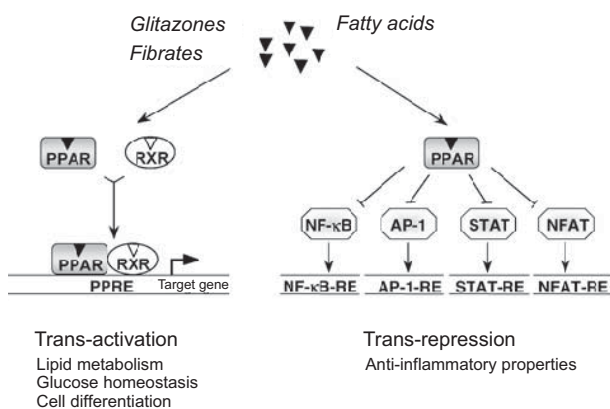


Fig. 3. Regulation of gene expression by PPARs. PPARs influence gene expression either by 'transactivation' through direct binding to PPREs or through indirect interference with other transcription factor pathways (AP-1, NF-κB, STAT and NFAT) leading in most cases to transcription inhibition ('transrepression'); (Reprinted with permission from Nature Publishing Group, [16])

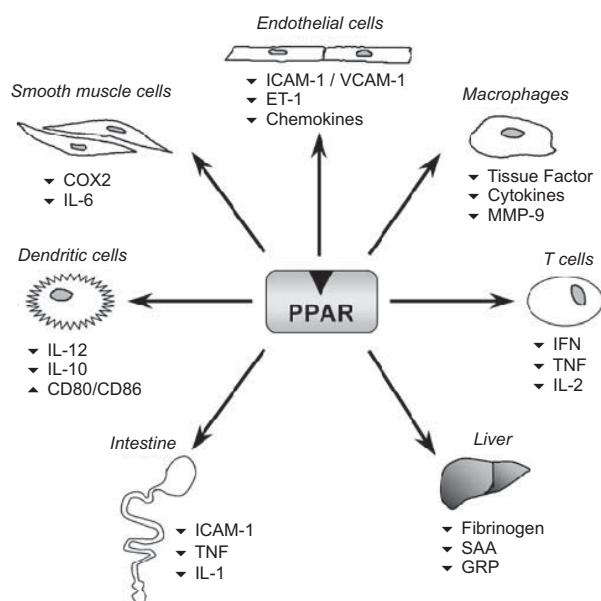


Fig. 4. Control of inflammation by PPARs. PPARs are modulators of the inflammatory response in different cell types and tissues (Reprinted with permission from Nature Publishing Group, [16])

has been suggested that stimulation of peroxisome proliferator-activated receptors (PPARs) and inhibition of nuclear factor kappa B (NF-κB) and other transcription factors play an important role in the mechanism of actions of NSAIDs.

PPARs are members of the nuclear receptor family. Three genes encode the peroxisome proliferator-activated receptor (PPAR) family members, PPAR-α, PPAR-β/δ and PPAR-γ. All three PPARs are widely expressed in different tissues. Their relative levels, however, differ between tissues reflecting their different biological functions. PPAR-α is highly expressed in the liver, skeletal muscle and heart. PPAR-γ is detected in adipose tissue, but is also present at high levels in the intestine, breast tissue, etc. PPAR-β/δ is ubiquitously expressed with the highest levels in the skin and skeletal muscle. Cells of the vascular wall and the immune system express all three PPARs. PPARs are activated by natural ligands such as fatty acids, eicosanoids and oxidized fatty acids and by pharmacological drugs. The lipid-lowering fibrates and the antidiabetic glitazones are synthetic ligands for PPAR-α and PPAR-γ, respectively. Influence of PPARs on gene expression may occur directly through promoter-binding and transcriptional modulation of target genes or through indirect interference with other transcription factor pathways leading in most cases to transcription inhibition (Fig. 3). PPARs regulate transcription of target genes involved in lipid and lipoprotein metabolism, glucose homeostasis and cell differentiation (particularly adipogenesis). PPARs inhibit the activation of certain inflammatory response genes acting as transrepressors.

PPARs and inflammation control

The first evidence for the role of PPARs in inflammation was obtained in PPAR-α-deficient mice which displayed a prolonged inflammation in response to the proinflammatory leukotriene B4 (LTB4). Recent studies aimed to delineate the cellular and molecular mechanisms by which PPAR-α modulates the inflammatory response. Activated PPAR-α blocks the production of inflammatory response markers, such as endothelin-1, intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1) in endothelial cells and tissue factor, matrix metalloproteinase (MMP)-9 and tumor necrosis factor alpha (TNF-α) in macrophages. In human aortic smooth muscle cells (SMC), PPAR-α activation by fibrates inhibits interleukin-1 (IL-1)-stimulated IL-6 secretion and 6-keto platelet growth factor (PGF)1 production by de-

creasing IL-6 and COX-2 gene transcription. Furthermore, aortic explants from PPAR- α -deficient mice showed an intensified response to inflammatory stimuli such as lipopolysaccharide (LPS), as indicated by increased IL-6 secretion. Taken together, these observations provide evidence that PPAR- α plays a role in the inflammatory response in different cell types (Fig. 4). Studies addressing the molecular mechanisms of the anti-inflammatory actions of PPARs suggest that many of the effects of PPARs are mediated by an inhibition of proinflammatory transcription pathways, such as NF- κ B, AP-1, NFAT, C/EBP or Smad3, *via* protein-protein interaction and competition for cofactor recruitment [16].

The NF- κ B family of transcription factors plays a major role in the regulation of the expression of a number of genes implicated in cell growth, inflammation, and apoptosis. The NF- κ B/Rel family is composed of five members, c-Rel, p65, Rel B, p50, and p52. In most non-activated cells, NF- κ B is in a cytoplasmic inactive complex through its association with the inhibitory proteins I κ Bs. Inducers of NF- κ B, which include inflammatory cytokines, reactive oxygen species, and viral products, activate a dimeric I κ B kinase (IKK) complex, which phosphorylates I κ B α on Ser-32 and Ser-36 leading to subsequent ubiquitination and degradation of I κ B α and release of NF- κ B proteins. Free NF- κ B dimers translocate to the nucleus where they regulate target gene transcription [19].

Use of NSAIDs

NSAIDs are used in the following diseases: rheumatoid arthritis, juvenile arthritis, ankylosing spondylitis, psoriatic arthritis, systemic lupus, crystal arthritis, posttraumatic pain, headache, toothache, upper respiratory tract infections, early stage cancer pain, urologic diseases, and tendency to thrombotic events [20, 76, 91, 104].

Side effects of NSAIDs

It is well-known that therapy with COX inhibitors is associated with a number of side effects including

gastrointestinal erosions, and renal and hepatic insufficiency. Such critical adverse reactions are highly dependent on COX-1 inhibition [12]. The most common side effect of NSAIDs is gastrointestinal (GI) toxicity. GI toxicity has been attributed to the inhibition of the COX-1-mediated generation of the cytoprotective prostanoids, such as prostaglandin PGE₂ and PGI₂ [52]. GI system damage may vary from hidden blood loss to ulcer perforations [7, 98]. NSAIDs may also cause a rise of serum creatine levels and induce hypercalcemia, interstitial nephritis, proteinuria, and acute renal dysfunction (renal toxicity) [66, 72, 76, 99]. Due to a reduced production of PGs, such as PGI₂, PGE₂ involved in the regulation of renal blood circulation, the rate of glomerular filtration is reduced. Especially in patients with reduced renal function, this results in retention of water, hypertension and, in some cases, to renal failure.

The inhibition of COX in thrombocytes leads to decreased production of thromboxane A₂. This phenomenon prolongs bleeding time and leads to inhibition of platelet aggregation [18]. In addition, NSAIDs may increase liver enzyme levels, and may cause agranulocytosis, aplastic anemia, toxic epidermal necrosis, cartilage metabolism dysfunction (NSAIDs block glucosaminoglycan synthesis), Reye's syndrome, vertigo [2, 8, 30, 31, 63, 65, 105]. Bronchoconstriction with asthmatic events is also a severe side effect of NSAIDs. The reduced amount of bronchodilating PGE₂ and a shift in the metabolic pathway from the COX pathway to the 5-lipoxygenase pathway are responsible for the bronchoconstricting effect of NSAIDs. The latter pathway metabolizes 'overflow' AA, which cannot be transformed by the blocked COX pathway. The final leukotrienes act as bronchoconstrictors [18].

Selective COX-1 and non-selective COX Inhibitors

These inhibitors include many groups (Tab. 1). We will discuss two groups of these inhibitory drugs in this review:

1. Salicylates (selective COX-1 inhibitors);
2. Phenylpropionic acid derivatives (non-selective COX inhibitors).

Salicylates

Acetylsalicylic acid (ASA), also known as aspirin, is the most commonly used drug amongst salicylates. It is an anti-inflammatory, analgesic, and antipyretic drug, which is not COX-2 specific and inhibits prostaglandin and thromboxane synthesis [53, 82]. ASA inhibits COX-1 more than COX-2 and the inhibition is irreversible [24]. ASA is effective in the treatment of inflammatory diseases, especially arthritis and carditis, and in acute rheumatoid fever [31, 92]. It is also administered in acute coronary heart diseases and chronic treatment of cerebral sinovenous thrombosis [3, 34]. ASA may induce epithelial damage in stomach mucosa, ulcer, and bleeding [37, 41, 71]. The bleeding arises from a reduction in synthesis of coagulation factors in the liver and inhibition of thrombocyte aggregation [51]. Allergic reactions, such as bronchospasm, vasomotor rhinitis, and angioedema may occur after ASA administration [2]. Salicylates have been found to have nephrotoxic and hepatotoxic effects [66]. Sodium salicylate has weaker analgesic, antipyretic, and anti-inflammatory effects in comparison with ASA. In addition, its antiaggregatory effect is not obvious, since it inhibits thromboxane synthase [31]. It is known to be as effective as ASA in rheumatoid diseases, although it is a weaker COX inhibitor than ASA. The fact that sodium salicylate does not cause gastric damage is in accordance with its ineffectiveness as COX-1 inhibitor [93]. Difluorophenyl derivative of salicylic acid, diflunisal, has anti-inflammatory, analgesic, and antipyretic effects. It weakly inhibits COX-1 and COX-2. Duration of diflunisal effect is longer than other salicylates. It is mainly used as an analgesic in muscle-skeleton system diseases and has side effects similar to salicylates [45]. Other salicylate derivatives are not used to produce systemic effects.

Phenylpropionic acid derivatives

This group contains analgesic, antipyretic and anti-inflammatory drugs, such as ibuprofen, naproxen, ketoprofen, fenbufen, tiaprofenic acid, fenprofen. These drugs inhibit both COX isoforms [11, 15, 35, 93]. Ibuprofen is effective in the treatment of rheumatoid arthritis, osteoarthritis, gout arthritis, and diseases with pain (headache, toothache, and dysmenorrhea) and reduces fever by inhibiting prostaglandin synthesis [1, 36, 45, 74]. It may cause GI side effects, renal dysfunction, meningeal syndrome, and bone marrow de-

pression [13, 39, 41, 43, 61, 77, 107]. Naproxen is an anti-inflammatory, analgesic and antipyretic drug with a long-term effect. It inhibits both COX isoforms [43]. It strongly inhibits functions of leukocytes at the inflammatory site [31, 45, 77] and it is frequently used in rheumatoid arthritis, ankylosing spondylitis, and traumatic damage to joints and surrounding tissues [69]. Most common side effects of naproxen are GI irritation [78], headache, vertigo, and depression [77, 97]. Ketoprofen effects are similar to naproxen. It has both central and peripheral analgesic effects [28]. Fenbufen and tiaprofen are anti-inflammatory, analgesic and antipyretic drugs. The former has fewer gastropathic effects, and the latter does not inhibit proteoglycan synthesis in cartilage tissues [46, 47].

Selective COX-2 inhibitors

Selective inhibitors of COX-2 are drugs whose therapeutic effects are as strong as conventional NSAIDs but which lead to fewer side effects [77].

Meloxicam, a relatively selective COX-2 inhibitor, has a more potent inhibitory effect on COX-2 than on COX-1 (Tab. 1) [44, 67]. However, COX-2 selectivity of meloxicam at high doses decreases and inhibition on COX-1 increases. Meloxicam is well tolerated by patients [27]. It has been as effective as other NSAIDs in the treatment of rheumatoid arthritis, osteoarthritis, and ankylosing spondylitis and was shown to have a better safety profile [40].

Nimesulide is a relatively selective COX-2 inhibitor and an analgesic, antipyretic and anti-inflammatory drug (Tab. 1) [35]. Studies have shown that nimesulide inhibits COX-2 *in vitro* up to five-fold more potently than COX-1 [17].

Nimesulide inhibits COX-2 more potently at lower doses, but at higher doses its COX-2 selectivity is abolished and it inhibits COX-1 more strongly than COX-2 [25]. Antipyretic effect of nimesulide is based on inhibition of prostaglandin synthesis [15].

Pain reducing effect of nimesulide has been found to arise, in part, from inhibition of cytokines [22], which are known to have a role in inflammatory process [85]. Nimesulide induces less damage to the gastric tissue. It did not induce gastric damage, even when given with steroidal anti-inflammatory drug, prednisolone [35]. Nimesulide also has antioxidant

properties [38]. It is used in the treatment of inflammatory pain [6]. Highly selective COX-2 inhibitory drugs such as celecoxib and rofecoxib have been developed since they have less adverse effects. They possess analgesic, antipyretic and anti-inflammatory effects as potent as traditional anti-inflammatory drugs [70]. Celecoxib and rofecoxib inhibit COX-2 375 and 800 times more strongly than COX-1, respectively [36]. It has been reported that these drugs do not induce damage to the stomach tissue [13]. Rofecoxib and celecoxib are as effective as other NSAIDs in inflammatory diseases [76], but recent studies have demonstrated that these drugs may result in thrombotic cardiovascular problems [58]. Therefore, the use of their (particularly rofecoxib and celecoxib) is controversial in recent days.

Recently some new coxibs (i.e. etoricoxib, valdecoxib, parecoxib and lumiracoxib) with increased biochemical COX-2 selectivity over that of rofecoxib and celecoxib have been developed. Valdecoxib has *in vitro* evidenced higher biochemical selectivity than celecoxib. This may be relevant to the improved gastrointestinal safety. Parecoxib, a prodrug of valdecoxib, is the only injectable selective COX-2 inhibitor. Etoricoxib displays slightly improved COX-2 selectivity than rofecoxib. Lumiracoxib, the most selective COX-2 inhibitor *in vitro*, is the only acidic coxib. These new molecules are suggested to have comparable effect to nonselective NSAIDs in the treatment of osteoarthritis, rheumatoid arthritis and acute pain, but they have similar renal adverse-effects, in several randomized clinical studies. The apparent dose-dependence of renal toxicity may limit the use of higher doses of the novel coxibs for improved effect [88].

As selective COX-2 inhibitors do not inhibit thromboxane A₂, incidence of bleeding is reduced. It has been hypothesized that these drugs cannot be used instead of aspirin [25]. The COX-2 isoform produces PGs at inflammatory sites as well as PGI₂, which is a vasodilator and inhibitor of platelet aggregation. Selective COX-2 inhibitors would have no effect on thromboxane A₂ production, but by decreasing PGI₂ production may disturb the natural balance between prothrombotic thromboxane A₂ and antithrombotic PGI₂, and may lead to an increase in thrombotic cardiovascular events [58]. Although these drugs are used in the treatment of rheumatoid arthritis, osteoarthritis, and inflammatory diseases [5, 6, 70, 79], they may cause blood pressure impairments and other car-

diorenal events in patients with hypertension. Some studies have shown that edema development and high diastolic blood pressure were observed in patients with hypertension taking rofecoxib and celecoxib. In celecoxib-administered patients, edema and blood pressure impairments were less frequent than in patients administered rofecoxib [102]. In another study, it was seen that rofecoxib enhanced markedly systolic blood pressure, while celecoxib did not increase it [103]. In addition, rofecoxib had a higher renal toxicity when compared to celecoxib and traditional NSAIDs [107].

There is apparent evidence that both COX-1 and COX-2 are present in the kidney in constitutive and inducible forms, which suggests that selective COX-2 inhibitors will have the same effects on renal prostaglandins as nonselective NSAIDs. The PGs that are most important in the kidney are PGE₂ and PGI₂, or prostacyclin. PGE₂ decreases sodium reabsorption at the thick ascending limb of the loop of Henle and PGI₂ stimulates renin release, which in turn increases aldosterone. Aldosterone increases sodium reabsorption and potassium secretion at the distal nephron. Prostacyclin is also a strong vasodilator that maintains glomerular filtration rate (GFR) and renal blood flow in patients with decreased actual or effective circulating volume. In healthy individuals, the vasodilatory role of PGI₂ is not operative and has little importance in renal hemodynamics. By understanding the physiological effects of renal PGs and their function, the consequences of inhibiting them can be predicted. Inhibiting PGE₂ will cause sodium retention, which can be manifested as weight gain, peripheral edema, or rarely, congestive heart failure. Blood pressure may also increase because of sodium retention. This occurs most often in patients with hypertension who are being treated with antihypertensive medications. In addition, inhibition of PGI₂ can cause hyperkalemia or, in patients at risk for adverse renal effects, acute renal failure [9]. Effects of selective COX-2 inhibitors on delayed wound healing (in gastric tissue) have been discussed in the following section.

Effects of some NSAIDs on gastric tissue

Effects of indomethacin, naproxen, ibuprofen, nimesulide, and diclofenac on rat stomach tissue have been

investigated. Indomethacin was observed to induce the largest damage to the gastric tissue. On the other hand, diclofenac induced the smallest lesions, naproxen and ibuprofen induced moderate damage to the gastric tissue, while nimesulide did not provoke ulcer [83]. The studies of Mitchell et al. [55] indicated that indomethacin and ibuprofen inhibited COX-1 more strongly than COX-2, while diclofenac inhibited COX-2 more than COX-1. These data show that the results are in accordance with the hypothesis of COX-1 and COX-2. In addition, ranitidine and nimesulide have been shown to completely prevent indomethacin-induced ulcers in rats [80]. Rofecoxib and celecoxib could not prevent indomethacin-induced ulcers [84]. In addition, both drugs enhanced stress-induced ulcers [68] while nimesulide, at a dose of 500 mg/kg, reduced stress-induced ulcers [83]. These data suggest that either COX hypothesis is not true or mechanisms of action of rofecoxib and celecoxib are not sufficiently defined. Prevention of indomethacin-induced ulcers by nimesulide suggested that there might be a chemical antagonism between indomethacin and nimesulide. Studies have emphasized that nimesulide and ranitidine do not chemically react with indomethacin [81], and there is no chemical antagonism between indomethacin and nimesulide. In a recent study by Kataoka et al. [35], it has been reported that indomethacin causes inhibition of COX-1 in gastric tissue, lowers PGE₂ levels, and increases acid secretion, myeloperoxidase (MPO) activity, stomach movement, while nimesulide relatively inhibits COX-2, does not affect PGE₂ and MPO levels and reduces gastric acid secretion. In another study, indomethacin increased hydrochloric acid (HCl) secretion and lowered bicarbonate (HCO₃) and PGE₂ synthesis [86].

There are several more issues that need to be addressed. May nimesulide be preventing indomethacin-induced ulcers by inhibiting H₂ receptors? Is COX-2 selectivity of nimesulide abolished at high doses? May ranitidine have anti-inflammatory effect?

Nimesulide and famotidine, when given together, inhibited HCl secretion more than when given alone, and this indicates that nimesulide does not affect histamine H₂ receptors [89]. Nimesulide, at a dose of 500 mg/kg, prevented stress-induced ulcers better than H₂ receptor blockers [83]. Both COX isoforms must be inhibited to provoke ulcer [62]. Should both enzymes be activated to induce gastroprotective effect?

These data demonstrate that mechanisms of action of drugs under discussion are multidirectional and complex. These drugs or their derivatives, which belong to the same group, have distinct pharmacological effects, side effects and potencies which imply that there may be more than two, five, or even tens of COX isoforms. The drugs that are well tolerated by some patients may cause serious adverse effects in other patients. Do effects of this kind arise from insufficiency of COX isoforms rather than from any organ pathology? Such questions and others will remain unanswered until new reports elucidate the unknown aspects of the relationship between COX isoforms and their physiological functions. Especially, the suggestion that COX may have a third isoform in the brain will be the focus of forthcoming studies. Until satisfactory data become available, clinicians must use NSAIDs cautiously.

References:

1. Amadio P: Peripherally acting analgesics. *Am J Med*, 1984, 10, 17–25.
2. Amadio P Jr, Cummings DM, Amadio P: Nonsteroidal anti-inflammatory drugs. *Postgrad Med*, 1993, 93, 73–76, 79–81, 85–88.
3. Aviles RJ, Bhatt DL: Antiplatelet therapies in combination for the treatment of patients with stable and unstable coronary artery disease. *J Thromb Thrombolysis*, 2002, 13, 177–182.
4. Bednarek D, Ciesielska AS, Zdisinska B, Kondracki M, Paduch R, Szerszen MK: The effect of steroidal and non-steroidal anti-inflammatory drugs on the cellular immunity of calves with experimentally induced local lung inflammation. *Vet Immunol Immunopathol*, 1999, 71, 1–15.
5. Bertolini A, Ottani A, Sandrini M: Dually acting anti-inflammatory drugs: a reappraisal. *Pharmacol Res*, 2001, 44, 437–450.
6. Bianchi M, Broggin M: Anti-hyperalgesic effects of nimesulide: studies in rats and humans. *Int J Clin Pract Suppl*, 2002, 128, 11–19.
7. Bjorkman DJ: Current status of nonsteroidal anti-inflammatory drugs (NSAID) use in the United States: risk factors and frequency of complications. *Am J Med*, 1999, 107, 3S–8S.
8. Bort R, Ponsoda X, Jover R, Lechon MJG, Castell JV: Diclofenac toxicity to hepatocytes: a role for drug metabolism in cell toxicity. *J Pharmacol Exp Ther*, 1999, 288, 65–72.
9. Brater DC: Renal effects of cyclooxygenase-2-selective inhibitors. *J Pain Symptom Manage*, 2002, 23, S15–S20.

10. Brooks PM, Day RO: Nonsteroidal anti-inflammatory drugs: differences and similarities. *N Eng J Med*, 1991, 324, 1716–1725.
11. Buritova J, Besson JM: Peripheral and/or central effects of racemic, S (+) and R (–)-flurbiprofen on inflammatory nociceptive processes: a c-fos protein study in the rat spinal cord. *Br J Pharmacol*, 1998, 125, 87–101.
12. Burdan F, Szumilo J, Klepacz R, Dudka J, Korobowicz A, Tokarska E, Cendrowska-Pinkosz M et al.: Gastrointestinal and hepatic toxicity of selective and non-selective cyclooxygenase-2 inhibitors in pregnant and non-pregnant rats. *Pharmacol Res*, 2004, 50, 533–543.
13. Buttgereit F, Burmester G, Simon LS: Gastrointestinal toxic side effects of non-steroidal anti-inflammatory drugs and cyclooxygenase-2-specific inhibitors. *Am J Med*, 2001, 110, 135–195.
14. Celotti F, Laufer S: Anti-inflammatory drugs: new multi-target compounds to face an old problem. The dual inhibition concept. *Pharmacol Res*, 2001, 43, 419–436.
15. Chandra J, Bhatnagar SK: Antipyretics in children. *Indian J Pediatr*, 2002, 69, 69–74.
16. Chinetti G, Fruchart JC, Staels B: Peroxisome proliferator-activated receptors and inflammation: from basic science to clinical applications. *Int J Obes*, 2003, 27, S41–S45.
17. Cullen L, Kelly L, Connor SO, Fitzgerald DJ: Selective cyclooxygenase-2 inhibition by nimesulide in man. *J Pharmacol Exp Ther*, 1998, 287, 578–582.
18. Dannhardt G, Kiefer W: Cyclooxygenase inhibitors – current status and future prospects. *Eur J Med Chem*, 2001, 36, 109–126.
19. Delerive P, Gervois P, Fruchart JC, Staels B: Induction of I κ B α expression as a mechanism contributing to the anti-inflammatory activities of peroxisome proliferator-activated receptor- α activators. *J Biol Chem*, 2000, 275, 36703–36707.
20. Dougados M, Gueguen A, Nakache JP, Velicitat P, Veys EM, Zeidler H, Calin A: Ankylosing spondylitis: what is the optimum duration of a clinical study? A one-year versus a 6-week nonsteroidal anti-inflammatory drug trial. *Rheumatology*, 1999, 38, 235–244.
21. Ferraz JG, Sharkey KA, Reuter BK, Asfaha SH, Tigley AW, Brown ML, McKnight W et al.: Induction of cyclooxygenase-1 and 2 in rat stomach during endotoxemia: role in resistance to damage. *Gastroenterology*, 1997, 113, 195–204.
22. Ferreira SH: Peripheral analgesic sites of action of anti-inflammatory drugs. *Int J Clin Pract, Suppl*, 2002, 128, 2–10.
23. Flower RJ, Vane JR: Inhibition of prostaglandin synthetase in brain explains the antipyretic activity of paracetamol (4-acetaminodiphenol). *Nature*, 1972, 240, 410–411.
24. Frolich JCA: Classification of NSAIDs according to the relative inhibition of cyclooxygenase isoenzymes. *Trends Pharmacol Sci*, 1997, 18, 30–34.
25. Halter F, Tarnawski AS, Schmassmann A, Peskar BM: Cyclooxygenase-2 implications on maintenance of gastric mucosal integrity and ulcer healing: controversial issues and perspectives. *Gut*, 2001, 49, 443–453.
26. Hawkey CJ: Nonsteroidal anti-inflammatory drug gastropathy. *Gastroenterology*, 2000, 119, 521–535.
27. Hawkey C, Kahan A, Steinbruck K, Alegre C, Baumelou E, Begaud B, Dequeker J et al.: Gastrointestinal tolerability of meloxicam compared to diclofenac in osteoarthritis patients. International MELISSA study group. Meloxicam large-scale international study safety assessment. *Br J Rheumatol*, 1998, 37, 937–945.
28. Herrero JF, Parrado A, Cervero F: Central and peripheral actions of the NSAID ketoprofen on spinal cord nociceptive reflexes. *Neuropharmacology*, 1997, 36, 1425–1431.
29. Higgs GA, Eakins KE, Mugridge KG, Moncada S, Vane JR: The effects of nonsteroidal anti-inflammatory drugs on leukocyte migration in carrageenin-induced inflammation. *Eur J Pharmacol*, 1980, 66, 81–86.
30. IAAAS. Risk of agranulocytosis and aplastic anemia. *JAMA*, 1986, 256, 1749–1757.
31. Insel PA: Analgesic-antipyretic and anti-inflammatory agents and drugs employed in the treatment of gout. In: Goodman & Gilman's *The Pharmacological Basis of Therapeutics*. Eds. Hardman JG, Limbird LE, McGraw Hill Press, New York, 1996, 617–658.
32. Isakson PC, Verburg KM, Maziasz TJ, Geis GS: Selective inhibitors of COX-2. *Gastroenterology*, 1999, 12, 169–177.
33. Jacobs JWG, Bijlsma JWJ: NSAIDs: a critical appraisal. *Neth J Med*, 1996, 51, 198–204.
34. Kao A, Dlugos D, Hunter JV, Mamula P, Thorarensen O: Anticoagulation therapy in cerebral sinovenous thrombosis and ulcerative colitis in children. *J Child Neurol*, 2002, 17, 479–482.
35. Kataoka H, Horie Y, Koyama R, Nakatsugi S, Furukawa M: Interaction between NSAIDs and steroid in rat stomach: safety of nimesulide as a preferential COX-2 inhibitor in the stomach. *Dig Dis Sci*, 2000, 45, 1366–1375.
36. Kayaalp O: *Medicinal Pharmacology from Rational Cure Aspect* (in Turkish). Feryal Press, Ankara, 2000, 1026.
37. Kelly JP, Kaufman DW, Jurgelon JM, Sheehan J, Koff RS, Shapiro S: Risk of aspirin-associated major upper-gastrointestinal bleeding with enteric-coated or buffered product. *Lancet*, 1996, 348, 1413–1416.
38. Kotsinas A, Gorgoulis V, Zacharatos P, Zioris H, Tripodiadis F, Donta I, Kyriakidis M et al.: Antioxidant agent nimesulide and beta blocker metoprolol do not exert protective effects against rat mitochondrial DNA alterations in adriamycin-induced cardiotoxicity. *Biochem Biophys Res Commun*, 1999, 254, 651–656.
39. Laine L, Harper S, Simon T, Bath R, Johanson J, Schwartz H, Stern S et al.: A randomized trial comparing the effect of rofecoxib, a cyclooxygenase 2-specific inhibitor, with that of ibuprofen on the gastroduodenal mucosa of patients with osteoarthritis. *Gastroenterology*, 1999, 117, 776–783.
40. Land B, Distel M, Bluhmki E: A double-blind randomized, placebo-controlled study of efficacy and tolerance of meloxicam treatment in patients with osteoarthritis of the knee. *Scand J Rheumatol*, 1998, 27, 32–37.
41. Lanza FL, Rack MF, Simon TJ, Quan H, Bolognese JA, Hoover ME, Wilson FR et al.: Specific inhibition of cyclooxygenase-2 with MK-0966 is associated with less

- gastrointestinal damage than either aspirin or ibuprofen. *Aliment Pharmacol Ther*, 1999, 13, 761–770.
42. Laudanno OM, Cesolari JA, Esnarriaga J, Rista L, Piombo G, Maglione C, Aramberry L et al.: Gastrointestinal damage induced by celecoxib and rofecoxib in rats. *Dig Dis Sci*, 2001, 46, 779–784.
 43. Laudanno OM, Cesolari JA, Esnarriaga J, San MP, Bedini OA: *In vivo* selectivity of nonsteroidal anti-inflammatory drugs and gastrointestinal ulcers in rats. *Dig Dis Sci*, 2000, 45, 1359–1365.
 44. Laurence DR, Bennett PN, Brown MJ: Inflammation, arthritis and non-steroidal anti-inflammatory drugs (NSAIDs). In: *Clinical Pharmacology*, Eds. Bennett PN, Brown MJ, Churchill Livingstone, Edinburgh, 1997, 249–266.
 45. Lee J, Katayama S: Inflammation and non-steroidal anti-inflammatory drugs. In: *Textbook of Pharmacology*. Ed. Smith CM, Reynaud AM, Saunders, New York, 1992, 401–436.
 46. Lewis JR: New antirheumatic agents. *JAMA*, 1977, 237, 1260–1261.
 47. Mahdy AM, Galley HF, Abdel-Wahed MA, el Korny KF, Sheta SA, Webster NR: Differential modulation of interleukin-6 and interleukin-10 by diclofenac in patients undergoing major surgery. *Br J Anaesth*, 2002, 88, 797–802.
 48. Mahmud T, Rafi SS, Scott DL, Wrigglesworth JM, Bjarnason I: Nonsteroidal anti-inflammatory drugs and uncoupling of mitochondrial oxidative phosphorylation. *Arthritis Rheum*, 1996, 39, 1998–2003.
 49. Maricic N, Ehrlich K, Gretzer B, Schuligoi R, Respondek M, Peskar BM: Selective cyclooxygenase-2 inhibitors aggravate ischemia-reperfusion injury in the rat stomach. *Br J Pharmacol*, 1999, 128, 1659–1666.
 50. Masferrer JL, Seibert K, Zweifel B, Needleman P: Endogenous glucocorticoids regulate an inducible cyclooxygenase enzyme. *Proc Natl Acad Sci*, 1992, 89, 3917–3921.
 51. Meade TW, Roderick PJ, Brennan PJ, Wilkes HC, Kelleher CC: Extra-cranial bleeding and other symptoms due to low dose aspirin and low intensity oral anticoagulation. *Thromb Haemost*, 1992, 68, 1–6.
 52. Meagher EA: Balancing gastroprotection and cardioprotection with selective cyclooxygenase-2 inhibitors: clinical implications. *Drug Saf*, 2003, 26, 913–924.
 53. Mehta P: Aspirin in the prophylaxis of coronary artery disease. *Curr Opin Cardiol*, 2002, 17, 552–558.
 54. Michaluart P, Masferrer JL, Carothers AM, Subbaramaiah K, Zweifel BS, Koboldt C, Mestre JR et al.: Inhibitory effects of caffeic acid on the activity and expression of cyclooxygenase-2 in human oral epithelial cells and in a rat model of inflammation. *Cancer Res*, 1999, 59, 2347–2352.
 55. Mitchell JA, Akarasereenont P, Thiemerman C, Flower RJ, Vane JR: Selectivity of non-steroidal anti-inflammatory drugs as inhibitors of constitutive and inducible cyclooxygenase. *Proc Natl Acad Sci USA*, 1994, 90, 11693–11697.
 56. Mitchell JA, Belvisi MG, Akarasereenont P, Robbins RA, Kwon OJ, Croxtall J, Barnes PJ et al.: Induction of cyclooxygenase-2 by cytokines in human pulmonary epithelial cells: regulation by dexamethasone. *Br J Pharmacol*, 1994, 113, 1008–1014.
 57. Mitchell JA, Warner TD: Cyclooxygenase-2: pharmacology, physiology, biochemistry and relevance to NSAID therapy. *Br J Pharmacol*, 1999, 128, 1121–1132.
 58. Mukherjee D: Selective cyclooxygenase-2 (COX-2) inhibitors and potential risk of cardiovascular events. *Biochem Pharmacol*, 2002, 63, 817–821.
 59. Muscara MN, Vergnolle N, Lovren F, Triggle CR, Elliott SN, Asfaha S, Wallace JL: Selective cyclooxygenase-2 inhibition with celecoxib elevates blood pressure and promotes leukocyte adherence. *Br J Pharmacol*, 2000, 129, 1423–1430.
 60. Patrignani P: Nonsteroidal anti-inflammatory drugs, COX-2 and colorectal cancer. *Toxicol Lett*, 2000, 112–113, 493–498.
 61. Payan DG, Shearn MA: Non-steroidal anti-inflammatory drugs; nonopioid analgesics, drugs used in gout. In: *Basics and Clinical Pharmacology*. Ed. Katzung BG, Appleton-Lange, East Norwalk, 1989, 431–450.
 62. Peskar BM, Maricic N, Gretzer B, Schuligoi R, Schmassmann A: Role of cyclooxygenase-2 in gastric mucosal defense. *Life Sci*, 2001, 69, 2993–3003.
 63. Pinsky PF, Hurwitz ES, Gtunn WJ: Reye's syndrome and aspirin. *JAMA*, 1999, 260, 657–661.
 64. Rainsford KD: Profile and mechanisms of gastrointestinal and other side effects of non-steroidal anti-inflammatory drugs (NSAIDs). *Am J Med*, 1999, 107, 27S–35S.
 65. Rainsford KD, Skerry TM, Chindemi P, Delaney K: Effects of NSAIDs meloxicam and indomethacin on cartilage proteoglycan synthesis and joint responses to calcium phosphate crystals in dogs. *Vet Res Commun*, 1999, 23, 101–113.
 66. Ruiz JG, Lowenthal DT: NSAIDs and nephrotoxicity in the elderly. *Geriatr Nephrol Urol*, 1999, 7, 51–57.
 67. Schattenkirchner M: Meloxicam: a selective COX-2 inhibitor nonsteroidal anti-inflammatory drug. *Expert Opin Invest Drugs*, 1997, 6, 321–334.
 68. Schmedtje JF, Jr Li YS, Liu WL, Dubois RN, Runge MS: Hypoxia induces cyclooxygenase-2 *via* the NF- κ B p65 transcription factor in human vascular endothelial cells. *J Biol Chem*, 1997, 272, 601–608.
 69. Schnitzer TJ, Kamin M, Olson WH: Tramadol allows reduction of naproxen dose among patients with naproxen-responsive osteoarthritis pain. *Arthritis Rheum*, 1999, 42, 1370–1377.
 70. Schnitzer TJ, Truitt K, Fleischmann R, Dalgin P, Block J, Zeng Q, Bolognese J et al.: The safety profile, tolerability, and effective dose range of rofecoxib in the treatment of rheumatoid arthritis. Phase II Rofecoxib Rheumatoid Arthritis Study Group. *Clin Ther*, 1999, 21, 1688–1702.
 71. Sevak R, Paul A, Goswami S, Santani D: Gastroprotective effect of beta (3) adrenoreceptor agonist ZD 7114 and CGP 12177A in rats. *Pharmacol Res*, 2002, 46, 351.
 72. Shah AA, Thjodleifsson B, Murray FE, Kay E, Barry M, Sigthorsson G, Gudjonsson H et al.: Selective inhibition of COX-2 in human is associated with less gastrointestinal injury: a comparison of nimesulide and naproxen. *Gut*, 2001, 48, 339–346.

73. Siegle I, Klein T, Backman JT, Saal JG, Nusing RM, Fritz P: Expression of cyclooxygenase-2 in human synovial tissue. *Arthritis Rheum*, 1998, 41, 122–129.
74. Sikes DH, Agrawal NM, Zhao WW, Kent JD, Recker D, Verburg KM: Incidence of gastroduodenal ulcers associated with valdecoxib compared with that of ibuprofen and diclofenac in patients with osteoarthritis. *Eur J Gastroenterol Hepatol*, 2002, 14, 1101–1111.
75. Simon LS: Role and regulation of cyclooxygenase-2 during inflammation. *Am J Med*, 1999, 106, 37S–42S.
76. Simon LS: COX-2 inhibitors. Are they nonsteroidal anti-inflammatory drugs with a better safety profile? *Gastroenterol Clin North Am*, 2001, 30, 1011–1025.
77. Simon LS, Milis JA: Non-steroidal anti-inflammatory drugs. *N Engl J Med*, 1980, 302, 1237–1243.
78. Simon LS, Weaver AL, Graham DY, Kivitz AJ, Lipsky PE, Hubbard RC, Isakson PC et al.: Anti-inflammatory and upper gastrointestinal effects of celecoxib in rheumatoid arthritis. A randomized controlled trial. *JAMA*, 1999, 282, 1921–1928.
79. Simon LS, Yocum D: New and future drug therapies for rheumatoid arthritis. *Rheumatology*, 2000, 39, 36–42.
80. Suleyman H, Akcay F, Altinkaynak K: The effect of nimesulide on the indomethacin- and ethanol-induced gastric ulcer in rats. *Pharmacol Res*, 2002, 45, 155–158.
81. Suleyman H, Altinkaynak K, Gocer F, Maras A, Akcay F, Onuk MD, Gepdiremen A: Effect of nimesulide on the indomethacin- and ibuprofen-induced ulcer in rat gastric tissue. *Pol J Pharmacol*, 2002, 54, 255–259.
82. Suleyman H, Demirezer LO, Kuruuzum A: Analgesic and antipyretic activities of *Rumex patientia* extract on mice and rabbits. *Pharmazie*, 2001, 56, 815–817.
83. Suleyman H, Demirezer LO, Kuruuzum A, Akcay F: Gastroprotective and antiulcerogenic effects of *Rumex patientia* L. extract. *Pharmazie*, 2002, 57, 204–205.
84. Suleyman H, Demirezer LO, Kuruuzum A: Effects of *Rumex patientia* root extract on indomethacin and ethanol-induced gastric damage in rats. *Pharmazie*, 2004, 59, 147–149.
85. Suto MJ, Ransone LJ: Novel approaches for the treatment of inflammatory diseases: inhibitors of NF-kappa B, and AP-1. *Curr Pharm Des*, 1997, 3, 515–528.
86. Takeuchi K, Kagawa S, Mimaki H, Aoi M, Kawauchi S: COX and NOS isoforms involved in acid-induced duodenal bicarbonate secretion in rats. *Dig Dis Sci*, 2002, 47, 2116–2124.
87. Tani S, Suzuki T, Kano S, Tanaka T, Sunaga K, Morishige R, Tsuda T: Mechanisms of gastric mucus secretion from cultured rat gastric epithelial cells induced by carbachol, cholecystokinin octapeptide, secretion, and prostaglandin E₂. *Biol Pharm Bull*, 2002, 25, 14–18.
88. Tacconelli S, Capone ML, Patrignani P: Clinical pharmacology of novel selective COX-2 inhibitors. *Curr Pharm Des*, 2004, 10, 589–601.
89. Tavares IA, Borelli F, Welch NJ: Inhibition of gastric acid secretion by nimesulide: a possible factor in its gastric tolerability. *Clin Exp Rheumatol*, 2001, 19, 13–15.
90. Tegeder I, Neupert W, Guhring H, Geisslinger G: Effects of selective and unselective cyclooxygenase inhibitors on prostanoic release from various rat organs. *J Pharmacol Exp Ther*, 2000, 292, 1161–1168.
91. ten Wolde S, Breedveld FC, Hermans J, Vandenbroucke JP, van de Laar MA, Markusse HM, Janssen M et al.: Randomised placebo-controlled study of stopping second-line drugs in rheumatoid arthritis. *Lancet*, 1996, 347, 347–352.
92. Thomas J, Straus WL, Bloom BS: Over-the-counter non-steroidal anti-inflammatory drugs and risk of gastrointestinal symptoms. *Am J Gastroenterol*, 2002, 97, 2215–2219.
93. Vane JR, Botting RM: Mechanism of action of anti-inflammatory drugs. *Scand J Rheumatol Suppl*, 1996, 102, 9–21.
94. Vane JR, Botting RM: Mechanism of action of nonsteroidal anti-inflammatory drugs. *Am J Med*, 1998, 104, 2S–8S.
95. Vane JR, Flower RJ, Botting RM: History of aspirin and its mechanism of action. *Stroke*, 1990, 21, Suppl 12, 12–23.
96. Vane JR, Mitchell JA, Appleton I, Tomlinson A, Bishop-Bailey D, Croxtall J, Willoughby DA: Inducible isoforms of cyclooxygenase and nitric oxide synthase in inflammation. *Proc Natl Acad Sci USA*, 1994, 91, 2046–2050.
97. Wallace CA: The use of methotrexate in childhood rheumatoid diseases. *Arthritis Rheum*, 1998, 41, 381–391.
98. Wallace J: Nonsteroidal anti-inflammatory drugs and gastroenteropathy: the second hundred years. *Gastroenterology*, 1997, 112, 1000–1016.
99. Wallace JL: Distribution and expression of cyclooxygenase isoenzymes, their physiological roles and the categorization of nonsteroidal anti-inflammatory drugs. *Am J Med*, 1999, 107, 1S–16S.
100. Wallace JL, Bak A, McKnight W, Asfaha S, Sharkey KA, MacNaughton WK: Cyclooxygenase-1 contributes to inflammatory responses in rats and mice: implications for gastrointestinal toxicity. *Gastroenterology*, 1998, 15, 101–109.
101. Warner TD, Giluliano F, Vojnovic I, Bukasa A, Mitchell JA, Vane JR: Nonsteroidal drug selectivities for cyclooxygenase-1 rather than cyclooxygenase-2 are associated with human gastrointestinal toxicity: a full *in vitro* analysis. *Proc Natl Acad Sci USA*, 1999, 96, 7563–7568.
102. Whelton A, Fort JG, Puma JA, Normandin D, Bello AE, Verburg KM: Cyclooxygenase-2 specific inhibitors and cardiorenal function: a randomized, controlled trial of celecoxib and rofecoxib in older hypertensive osteoarthritis patients. *Am J Ther*, 2001, 8, 85–95.
103. White WB, Whelton A, Fort JG: Rofecoxib but not celecoxib increases systolic blood pressure in hypertensive patients treated with ACE inhibitors and beta-blockers. *College Am J Cardiol*, 2002, 39, Suppl, 249A.
104. Williams HJ, Ward JR, Egger MJ, Neuner R, Brooks RH, Clegg DO, Field EH et al.: Comparison of naproxen and acetaminophen in a two-year study of treatment of osteoarthritis of the knee. *Arthritis Rheum*, 1993, 36, 1196–1206.
105. Willoughby DA, Moore AR, Colville-Nash PR: COX-1, COX-2 and COX-3 and the future treatment of chronic inflammatory disease. *Lancet*, 2000, 355, 646–648.
106. Xie W, Chipman JG, Robertson DL, Erikson RL, Simmons DL: Expression of a mitogen-responsive gene en-

coding prostaglandin synthase is regulated by mRNA splicing. *Proc Natl Acad Sci USA*, 1991, 88, 2692–2696.

107. Zhao SZ, Reynolds MW, Lejkowith J, Whelton A, Arellano FM: A comparison of renal-related adverse drug reactions between rofecoxib and celecoxib, based on the

World Health Organisation/Uppsala Monitoring Centre safety database. *Clin Ther*, 2001, 23, 1478–1491.

Received:

April 5, 2004; in revised form June 13, 2007.